Water Treatment Methods for Control and Management of Algae

Everett J. Nichols, Ph.D., MSPH Scientific Director of Biopolymer Research



Presentation Overview

- 1. Algae & Cyanobacteria Classification
- 2. Nutrient Requirements and Assimilation
- 3. Algae Control Measures
- 4. Flocculation with the Biopolymer Chitosan
- 5. Phosphates
- 6. Phosphate Control Measures
- 7. Summary



Algae & Cyanobacteria Waterblooms

Green, red or brown colored water resulting from high density growth of algae or cyanobacteria

Marine Water

Red & brown tide coastal water & estuaries worldwide. Chesapeake Bay, North Carolina, Gulf of Mexico, Washington & Oregon Coast (red & green tides)

Freshwater

lakes, reservoirs, ponds (green scum) common indicator of eutrophication swimming pools, fountains, etc.



Algae & Cyanobacteria

<u>Algae</u>

Kingdom – Protista

- Eukaryotic (cell structure similar to multicellular plants and animals – contain cell nucleus, cytoplasmic organelles such as mitochondria, chloroplasts)
- Complex tissue development or multicellular reproductive structures lacking
- Oxygenic photosynthesis chlorophyll a
- Incapable of fixing nitrogen
- Some toxin producing (marine dinoflagellates) neurotoxins and hepatotoxins



Algae & Cyanobacteria

Cyanobacteria

Kingdom – Monera

- prokaryotic (no cell nucleus or cytoplasmic organelles such as mitochondria, chloroplasts)
- Simple single cell structure (rods, cocci, spirals and nonbranching filaments)
- Oxygenic photosynthesis chlorophyll a
- Accessory pigments –phycobiliproteins
- Capable of fixing atmospheric nitrogen
- Some toxin producing (similar to marine algae dinoflagellate toxins)
 - Gelatinous toxin released upon killing by chlorination or copper sulfate



Major Algae Groups

Common Name	# Species	Common Habitat
Green Algae	~7,000	Fresh water, salt water, damp soil
Yellow Green Algae	~600	Freshwater, salt water
Red Algae	~4,000	Saltwater, some freshwater
Brown Algae	~1,500	Saltwater, seaweeds, kelp
Golden Brown Algae (Diatoms)	~6,000	Freshwater, saltwater, soil
Dinoflagellates	~2,000	Saltwater, freshwater (red tides,fish kill)
Euglenoids	~6,000	Freshwater



Major Cyanobacterial Groups

Order	# Genera	Heterocysts	General Shape
Chroococcales	5	_	Unicellular, rods,/cocci, nonfilamentous aggregates
Pleurocapsales	3	_	Unicellular, rods,/cocci, nonfilamentous aggregates
Oscillatoriales	4	-	Filamentous, unbranched trichomes containing vegetative cells
Nostocales	6	+	Filamentous, unbranched trichomes may contain specialized cells
Stigonematales	3	+	Filamentous trichomes with either branches or more than one row of cells

From: Prescott, L.M., Harley, J.P., Klein, D.A. Microbiology, *Third Edition*, Wm. C. Brown Publishers, Dubuque, IA. 1996.



Algae & Cyanobacteria

Photosynthetic

 $\mathbf{CO}_2 + \mathbf{H}_2\mathbf{O} \xrightarrow{light} (\mathbf{CH}_2\mathbf{O}) + \mathbf{O}_2$

Cyanobacteria

Fix Nitrogen

 $\begin{array}{rl} Nitrogenase \\ N_2 + 8 e^- + 16 \text{ ATP} + 8 \text{ H}^+ &\longrightarrow & 2 \text{ NH}_3 + \text{ H}_2 + & 16 \text{ ADP} + & 16 \text{ P}_i \end{array}$



Nutrient Requirements of Algae & Cyanobacteria

Elements for Growth

 <u>Carbon</u>, hydrogen, <u>nitrogen</u>, oxygen, <u>phosphorus</u>, <u>sulfur</u>, calcium, potassium, magnesium, manganese, molybdenum, copper, iron, zinc, silicon, sodium, boron



Nitrogen, Phosphorus and Sulfur

- 1. Need to incorporate large quantities
 - Acquired from same source of carbon nutrients
 - Inorganic sources
- 2. Nitrogen
 - Synthesis of amino acids, purines, pyrimidines, enzyme cofactors
 - Nitrate reduction to ammonia and assimilation via glutamate dehydrogenase, glutamate synthase etc.
 - Cyanobacteria reduce atmospheric N₂ and assimilate via nitrogenase system
- 3. Sulfur
 - Synthesis of sulfhydryl-containing amino acids, enzyme cofactors
- 4. Phosphorus
 - Energy transfer (ATP), DNA, RNA (protein synthesis), membrane lipids



Phosphorus – Primary Growth Limiting Nutrient

- Published studies strongly support phosphate as the growth limiting nutrient for algae & cyanobacteria growth
- Schindler, D.W. *et al. (2008)* Eutrophication of lakes cannot be controlled by reducing nitrogen input: Results of a 37-year whole-ecosystem experiment. PNAS 105 (32): 11254-11258.

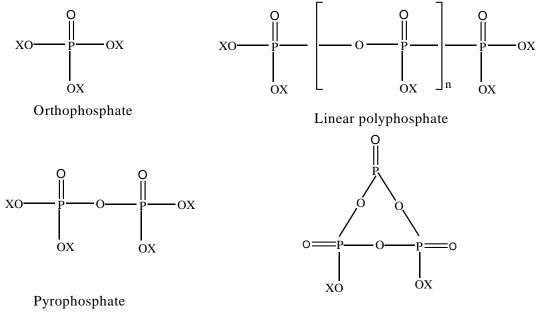


Phosphorus

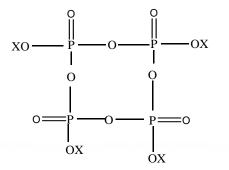
- Inorganic forms
 - ✓ Igneous rocks (calcium phosphates)
 - ✓ phosphate salts and esters of phosphoric acids
- Organic forms
 - contain 1-3 organic groups in ester linkage to oxygen
 - ATP, DNA, RNA, phospholipids, inositol phosphates, all formed by biological processes from orthophosphates

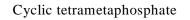


Examples of Inorganic Phosphates



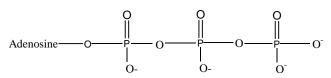
Cyclic trimetaphosphates



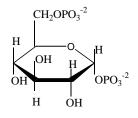




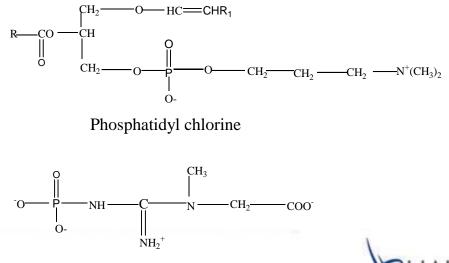
Examples of Organic Phosphates



Adenosine triphosphate (ATP)



Glucose 1,6-diphosphate





Creatine phosphate

Control Measures for Algae & Cyanobacteria

1. Chemical Disinfection

- Halogens (chlorine, bromine)
- Ionic silver chelates
- Copper sulfate
- Quaternary ammonium compounds
- 2. Ultraviolet Light
- 3. Ozone
- 4. Ultrasound
- 5. Aeration (dissolved oxygen)
- 6. Nutrient Control
 - Bioremediation
 - Nutrient Stripping

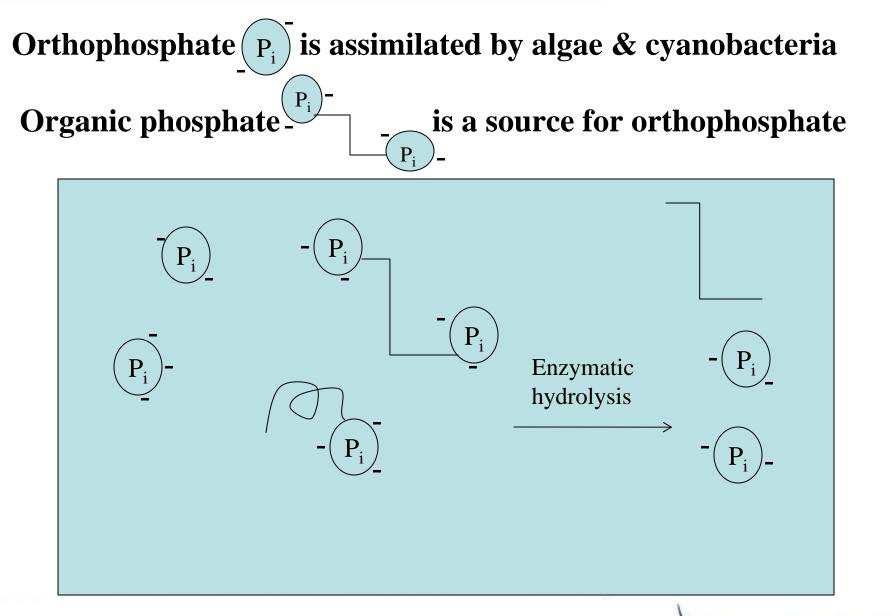


Nutrient Stripping

Reduce Phosphates (growth limiting nutrient)

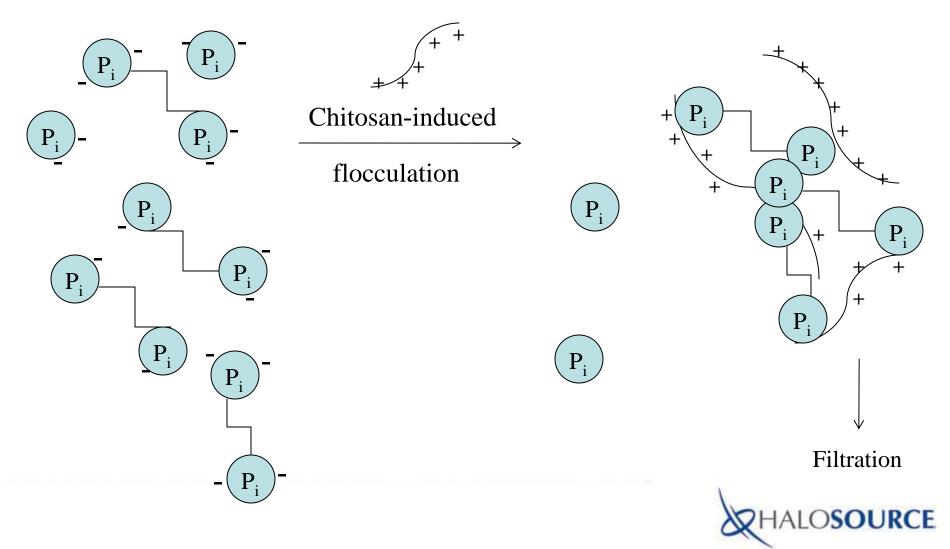
- Flocculation & filtration of <u>organic</u> phosphates
- Precipitation & filtration of <u>inorganic</u> phosphates



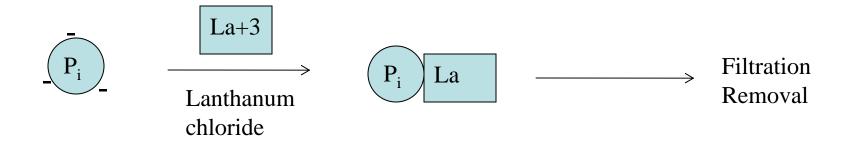




Removal of Organic Phosphates by Flocculation & Filtration



Removal of Inorganic Phosphate (Orthophosphate) by Precipitation & Filtration



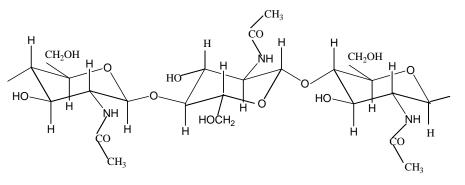


Natural Flocculant Biopolymer Chitosan

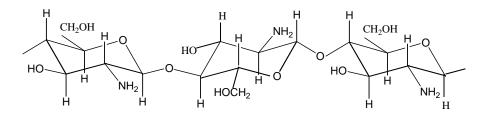
- Derived from chitin (structural polysaccharide of exoskeletons of crustaceans, insects, fungi)
- Structurally related to cellulose
- Cationic polysaccharide
- Biodegradable
- Binds to anionic suspended solids in water



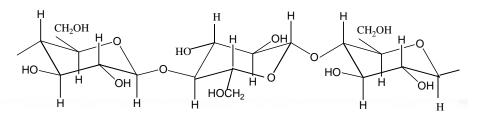
Structural Comparison of Chitosan



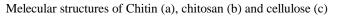
(a) CHITIN



(b) CHITOSAN

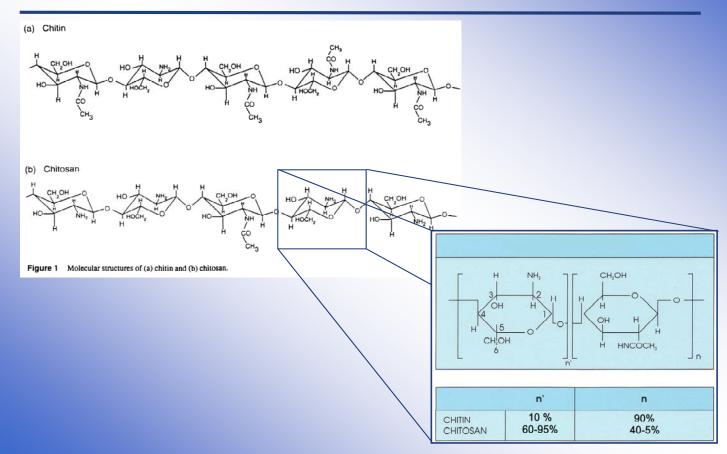


(C) CELLULOSE





Chitin/Chitosan Structure





N-Halochitosan - a patent protected derivative of chitosan flocs non-polar organics such as oils

N-Halochitosan is formed in the presence of halogens such as chlorine (hypochlorite and hypochlorous acid)

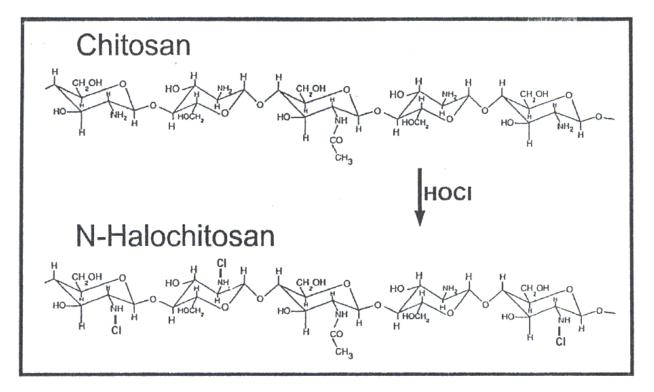
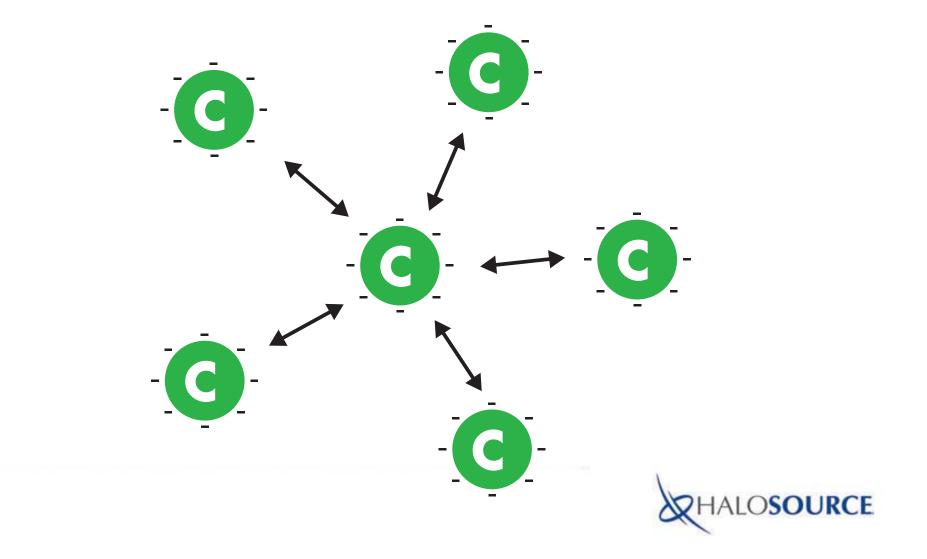


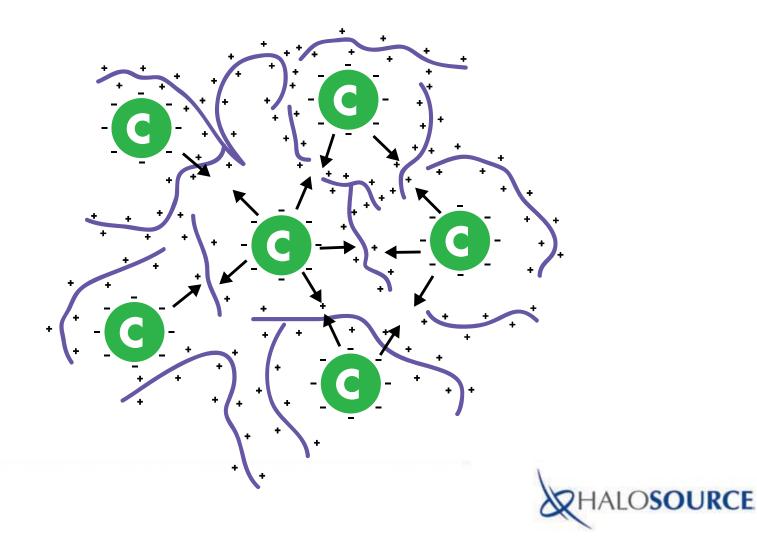
Figure 5 – Molecular Structure of N-Halochitosan



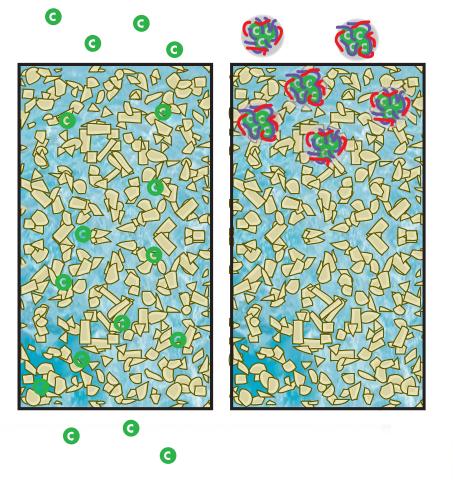
Colloid suspensions: surface charges cause mutual repulsion



Chitosan (cationic polymer) : charge neutralization reduces repulsion and suspension becomes unstable to form flocs

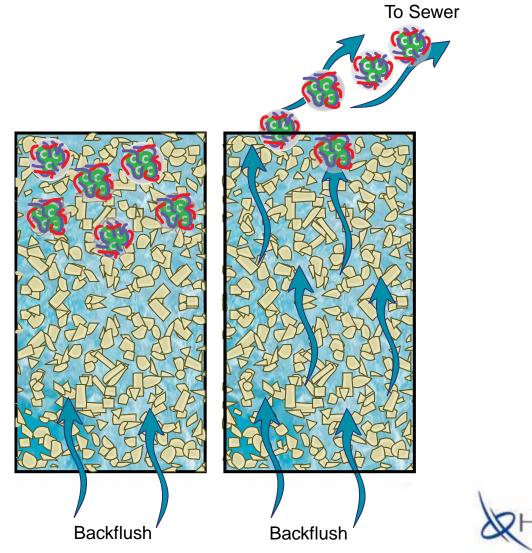


Sand bed filters can trap stable, firm floccules, and remove flocced sediment or colloidal insoluble organic phosphates from the flow of water





Backwash flushes trapped floccules into sanitary sewer system





Chitosan-Mediated Flocculation of Sediment



Suspended sediment-untreated

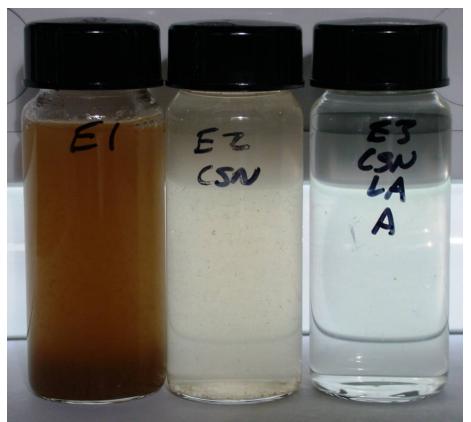
Seconds following addition of chitosan

Suspended sediment-untreated

Minutes following addition of chitosan



Treatment of Fish Waste with Chitosan and Lanthanum Chloride



No treatment E1

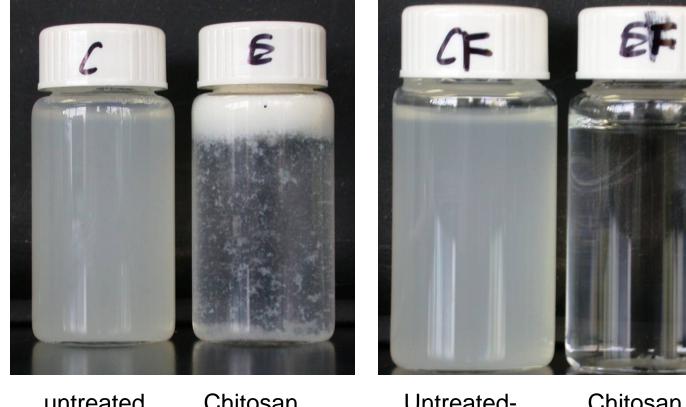
Chitosan treatment E2 Chitosan & Lanthanum chloride treatment E3 E1 (supernatant of diluted fish waste) Total phosphorus-572 ppm

E2 (E1 was treated w/ chitosan, allowed to settle and supernatant tested) Total phosphorus-142 ppm)

E3 (E2 was further treated w/lanthanum chloride and supernatant tested) Total phosphorus-2 ppm)



Flocculation & Filtration of Organic Phospholipid w/Chitosan



CF-total Phosphorus-33 ppm

CF-total Phosphorus-6.0 ppm

untreated

Chitosan treated

Untreatedsand filtered Chitosan treated & sand filtered HALOSOURCE

Flocculation & Filtration of Green Algae w/Chitosan



Algae nontreated Algae chitosantreated

Algae nontreated & sand filtered Algae chitosantreated & sand filtered

OSOURCE

Chemical Treatment Options for Orthophosphate Removal

Alum - $AI_2(SO_4)_3$ – most common

 $\begin{array}{rcl} & & & & & & & & \\ Al_2(SO_4)_3 \,+\, 6\,H_2O & & & & & \\ HCO_3^- &+\, H^+ & \longrightarrow & H_2CO_3 & \longrightarrow & CO_2 \,+\, H_2O \\ Al_2(SO_4)_3 \,+\, 3\,Ca(HCO_3)_2 & & & & & \\ Al_2(SO_4)_3 \,+\, 3\,Ca(HCO_3)_2 & & & & & \\ \end{array}$



Removal of orthophosphate w/ alum and ferric sulfate is a sorption process

 $AI(OH)_3$ sorbs orthophosphates \longrightarrow $AI(OH)_3 \cdot H_2PO4$

Advantages

• Low cost

Disadvantages

- Reduction of alkalinity (acid neutralization of HCO_{3⁻})
- Gelatinous precipitates
- Large quantities required to reduce PO₄-3 to low levels
- Toxicity concerns have been raised (AI+3)
- Potential for staining (iron)
- Increased CO₂ photosynthesis & algae growth



Lanthanides (trivalent rare earth metals)

- 1. Anion Substitution/Ion Exchange Lanthanum carbonate
- 2. Direct Precipitation Lanthanum chloride



Anion Substitution - exchange of carbonate for phosphate Lanthanum carbonate $La_2(CO_3)_3$ (insoluble)

 $\begin{array}{rcl} \text{La}_{2}(\text{CO}_{3})_{3} \ + \ 2 \ \text{PO}_{4}^{-3} & \xrightarrow{Slow} & 2 \ \text{La}\text{PO}_{4}^{-3} \ + \ 3 \ \text{CO}_{3}^{-2} \\ \text{CO}_{3}^{-2} \ + \ \text{H}_{2}\text{O} & \longrightarrow & \text{HCO}_{3}^{-3} \ + \ \text{OH}^{-} \\ \text{Ca}^{+3} \ + \ \text{CO}_{3}^{-2} & \longrightarrow & \text{CaCO}_{3} \end{array}$

Direct Precipitation of Orthophosphate Lanthanum chloride LaCl₃ (soluble)

$$La^{+3} + 3 Cl^{-} + PO_4^{-3} \xrightarrow{VeryFast} La PO_4 + 3 Cl^{-}$$



Advantages of LaCl₃ Compared to La₂(CO₃)₃

- Faster reaction kinetics
- Soluble reactant vs insoluble reactant
- No contributions of carbonate & potential CO₂
- Low potential for formation of scale
- Low potential for decreased hardness



Summary

- Algae and Cyanobacteria both contribute to water blooms.
- Cyanobacteria capable of fixing atmospheric nitrogen.
- Phosphorus is the key growth limiting nutrient.
- Important to remove <u>both</u> organic and inorganic forms of phosphate
- Phosphorus removal accomplished by chitosanmediated flocculation of organic phosphate and direct precipitation of orthophosphate by lanthanum chloride followed by filtration.



Acknowledgements

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Questions

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- Everett Nichols

